

**Table of Loci for Traits in Grapevine Relevant for Breeding and Genetics:**

Associated markers, their chromosomal localisation, and the donor genotype/species are given. Chromosomal position of a trait/allele is given in megabases according to the 12 x genome sequence of PN40024 (<http://www.genoscope.cns.fr/vitis>).

**Update: February 16, 2024**

The symbols were discussed and assigned at the International Conference on Grapevine Breeding and Genetics at Geneva, August 1 - 5, 2010. Follow up information on naming of loci will be provided on VIVC to avoid homonyms.

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Trait	Symbol	Chr	Position on chr [Mb]	Associated / flanking marker	Parent 1	Parent 2	Population size	Genotype of origin	Original species of trait	Reference	Comment
Resistance											
<i>Agrobacterium spec.</i> [crown gall]	<i>Rcg1</i>	15	7.1 9.3	UDV015 9M3-3	Kunbarát	Sárféhér	272	Kunbarát	<i>V. amurensis</i>	Kuczmog et al. (2012)	crown gall
<i>Botrytis cinerea</i> [bunch rot; gray mold]		2	5.0 5.0	VMC6F1 VMC3B10	Norton	Cabernet Sauvignon	182	Norton	<i>V. aestivalis</i>	Sapkota et al. (2019)	
<i>Botrytis cinerea</i> [leaf; gray mold]		2 7 2 7		RG ZSX	VS VT		177 176			Su et al. (2023)	leaf no marker data given
<i>Colletotrichum gloeosporioides</i> ( <i>Glomerella cingulata</i> ) [ripe rot]	<i>CgR1</i>	14	4.1	np19345	Cabernet Sauvignon	Shuang Hong	91	Shuang Hong	<i>V. amurensis</i>	Fu et al. (2019)	ripe rot
<i>Coniothyrium diplodiella</i> [white rot]	<i>Rcd1</i>	14	3.5 6.6	chr14_3541187 chr14_6602952	Zhuosexiang	Victoria	177	Zhuosexiang		Su et al. (2021)	white rot
<i>Daktulosphaira vitifoliae</i> [phylloxera]	<i>Rdv1</i>	13	GF13-1 21.9 GF13-1 21.5	GF.V3125 GF13-9 GF13-1 GF13-7	Börner		188	Börner	<i>V. cinerea</i>	Zhang et al. (2009)	
<i>Daktulosphaira vitifoliae</i>	<i>Rdv2</i>	14	4.9	S14_4921219	<i>V. cinerea</i> C2-50	Riesling	90	<i>V. cinerea</i> C2-50	<i>V. cinerea</i>	Smith et al. (2018)	root resistance
<i>Daktulosphaira vitifoliae</i>	<i>Rdv3</i>	14	5.0 4.8	S14_5049399 14_4805213	MN1264	MN1246	125 1023	MN1264 Seyval Blanc		Clark et al. (2018) Yin et al. (2022)	foliar resistance MN1264 = (MN1069 x Seyval Blanc)
<i>Daktulosphaira vitifoliae</i>	<i>Rdv4</i>	4			MN1264	MN1246	125	MN1264		Clark et al. (2018)	foliar resistance
<i>Daktulosphaira vitifoliae</i>	<i>Rdv5</i>	5			MN1264	MN1246	125			Clark et al. (2018)	root resistance
<i>Daktulosphaira vitifoliae</i>	<i>Rdv6</i>	7			VRH8771	Cabernet Sauvignon	135	VRH8771	<i>M. rotundifolia</i>	Rubio et al. (2020)	root resistance
<i>Daktulosphaira vitifoliae</i>	<i>Rdv7</i>	3	5.5	3_5494608	VRH8771	Cabernet Sauvignon	135	VRH8771	<i>M. rotundifolia</i>	Rubio et al. (2020)	root resistance
<i>Daktulosphaira vitifoliae</i>	<i>Rdv8</i>	10			VRH8771	Cabernet Sauvignon	135	VRH8771	<i>M. rotundifolia</i>	Rubio et al. (2020)	root resistance
<i>Diaporthe ampelina</i> ( <i>Phomopsis viticola</i> )	<i>Rda1</i>	15	19.6 S15_19591538 19.3	S15_19560016 S15_19591538 S15_19300044	Chardonnay Horizon Horizon	<i>V. cinerea</i> B9 <i>V. cinerea</i> B9 Illinois 547-1	148 162 366	<i>V. cinerea</i> B9 <i>V. cinerea</i> B9 Illinois 547-1	<i>V. cinerea</i>	Barba et al. (2018)	cane, cluster cane, cluster cane
<i>Diaporthe ampelina</i> ( <i>Phomopsis viticola</i> )	<i>Rda2</i>	7	1.2 1.8 3.1 3.1 1.9	VVMD7 VtZAG62 VvIB22 S7_3127568 S7_1912889	Horizon	<i>V. cinerea</i> B9	162	Horizon		Barba et al. (2018)	cane, cluster cane
<i>Elsinoë ampelina</i>	<i>Rea1</i>									Modesto et al. (in preparation)	
<i>Erysiphe necator</i>	<i>Ren1</i>	13		UDV020 18.4 18.4	Nimrang	Kishmish vatkana	310	Kishmish vatkana	<i>V. vinifera</i>	Hoffmann et al. (2008)	
<i>Erysiphe necator</i>	<i>Ren1.2</i>	13	17.9 18.2	SNP_13_17909186 SNP_13_18213673	Shavtsitska	Glera	184	Shavtsitska	<i>V. vinifera</i>	Possamai et al. (2021)	
<i>Erysiphe necator</i>	<i>Ren2</i>	14	26.9	CS25	Horizon	Illinois 547-1	58	Illinois 547-1		Dalbo et al. (2001)	
<i>Erysiphe necator</i>	<i>Ren3</i>	15	7.1 10.9 4.9 10.9 9.3	UDV015b VVIV67 ScORA7-760 VChr15CenGen02 GF15-28 / VVIV67 GF15-42	Regent	Lemberger	153	Regent		Welter et al. (2007) Akkurt et al. (2007) van Heerden et al. (2014) Zyprian et al. (2016) Zendler et al. (2017)	

Trait	Symbol	Chr	Position on chr [Mb]	Associated / flanking marker	Parent 1	Parent 2	Population size	Genotype of origin	Original species of trait	Reference	Comment		
<i>Erysiphe necator</i>	<i>Ren4</i>	18	26.9	VMC7f2	C166-043	F8909-08	42	C166-043	<i>V. romane</i> tii	Riaz et al. (2012)			
			26.9	SNPs	C87-41	B70-57	57	C87-41	<i>V. romane</i> tii	Mahanil et al. (2012)			
<i>Erysiphe necator</i>	<i>Ren5</i>	14	4.8	VMC9c1	Regale	Regale	191	Regale	<i>M. rotundifolia</i>	Blanc et al. (2012)			
			8.6	PN9-057	F2-35	V. piasezkii (DVIT2027)	277	V. piasezkii (DVIT2027)	<i>V. piasezkii</i>	Pap et al. (2016)			
<i>Erysiphe necator</i>	<i>Ren6</i>	9		9.1	PN9-068								
		0.2	VVIp17.1	F2-35	V. piasezkii (DVIT2027)	277	V. piasezkii (DVIT2027)	<i>V. piasezkii</i>	Pap et al. (2016)				
<i>Erysiphe necator</i>	<i>Ren7</i>	19	0.9	VMC9a2.1	GF.GA-47-42	Villard blanc	151			Zyprian et al. (2016)			
			13.2	UDV117									
<i>Erysiphe necator</i>	<i>Ren8</i>	18	1.4	CenGen6	Regent	Lemberger	153	Regent		Zendler et al. (2017)			
				SPS_P_SNP632GF									
<i>Erysiphe necator</i>	<i>Ren9</i>	15	1.4		Regent	Lemberger	153	Regent		Zendler et al. (2017)			
<i>Erysiphe necator</i>	<i>Ren10</i>	2	17.9	S2_17854965	MN1264	MN1214	147	Seyval blanc		Teh et al. (2017)			
				Haploblock validation	MN1264	MN1246	125						
<i>Erysiphe necator</i>	<i>Ren11</i>	15	13.7	rh_chr15_13698923	B37-28	C56-11	244	Tamiami	<i>V. aestivalis</i>	Karn et al. (2021)	B37-28 (Tamiami x <i>V. vinifera</i> )		
			15.3	rh_vhr15_15294725	Tamiami	M13	300						
<i>Erysiphe necator</i>	<i>Ren12</i>	13	22.8	13_22768514	<i>V. amurensis</i>	Valley Pearl	248	<i>V. amurensis</i>	<i>V. amurensis</i>	Sapkota et al. (2023)			
			27.0	13_26962710									
<i>Erysiphe necator</i>	<i>Ren13</i>										Cadle-Davidson et al. (in preparation)		
<i>Erysiphe necator</i>	<i>Ren14</i>										De Lorenzis et al. (in preparation)		
<i>Erysiphe necator</i>	<i>Ren15</i>										De Lorenzis et al. (in preparation)		
<i>Erysiphe necator</i>	<i>Ren16</i>										Hwang et al. (in preparation)		
<i>Erysiphe (Uncinula) necator</i>	<i>Run1</i>	12	13.1	VMC4f3.1	VRH3082-1-42	Cabernet Sauvignon	161	VRH3082-1-42	<i>M. rotundifolia</i>	Barker et al. (2005)	powdery mildew resistance originating from <i>Muscadinia</i> was named <i>Run</i>		
			20.4	VMC8g9									
			16.4	49MRP1.P2	VRH3082-1-42	Cabernet Sauvignon	2575			Feechan et al. (2013)			
			16.8	CB53.54		VRH3176-21-11	722						
						VRH3161-6-4	110						
						BC1.M. rotundifolia	139						
<i>Erysiphe (Uncinula) necator</i>	<i>Run2.1</i>	18	26.9	VMC7f2	JB81-107-11	Chenin Blanc	97	<i>Magnolia</i>	<i>M. rotundifolia</i>	Riaz et al. (2011)	resistant tissue: Cane rachis rachis fruit leaf, cane, rachis, fruit		
			20.9	VMCNgl3e3									
			23.4	VVIIn16	JB81-107-11	Tokay	47						
			26.9	VMC7f2									
			26.9	VMC7f2	A90-71	Flame Seedless	80						
<i>Erysiphe (Uncinula) necator</i>	<i>Run2.2</i>	18	26.9	VMC7f2	e2-9	Malaga Rosada	255	Trayshed	<i>M. rotundifolia</i>	Riaz et al. (2011)			
<i>Erysiphe necator</i>	<i>Sen1</i>	9	13.6 - 18.0	S8_19258484	<i>V. rupestris</i> B38	Chardonnay	85	Chardonnay	<i>V. vinifera</i>	Barba et al. (2014)			
<i>Grapevine fanleaf virus (GFVV)</i>	<i>rgflv1</i>	1	0.6	VMC4f8	(Riesling x	self-pollinated	87	Riesling	<i>V. vinifera</i>	Djennane et al. (2021)			
			1.5	Chr1_1535	Gewurztraminer)								
<i>Guignardia bidwellii</i>	<i>Rgb1</i>	14	26.7	GF14-42	GF.V3125	Börner	202	Börner		Rex et al. (2014)			
<i>Guignardia bidwellii</i>	<i>Rgb2</i>	16	15.3	VChr16c	GF.V312	Börner	202	Börner		Rex et al. (2014)			
<i>Meloidogyne javanica</i> (root knot nematode)	<i>MjR1</i>	18	31.2	S18_31160355	C2-50	Riesling	90	C2-50	<i>V. cinerea</i>	Smith et al. (2018)			
			34.0	S18_33954011									
Pierce's disease ( <i>Xylella fastidiosa</i> )	<i>Pdr1</i>	14	25.3	VMCNgl3h8	<i>V. rupestris</i>	<i>V. arizonica</i>	181		<i>V. arizonica</i>	Riaz et al. (2006)			
			26.6	VVIIn64						Riaz et al. (2008)			
			26.1	UDV095									
<i>Plasmopara viticola</i>	<i>Rpv1</i>	12	10.3	VVIb32	Syrah	28-8-78		28-8-78	<i>M. rotundifolia</i>	Merdinoglu et al. (2003)			
<i>Plasmopara viticola</i>	<i>Rpv2</i>	18			Cabernet Sauvignon	8624	129	8624	<i>M. rotundifolia</i>	Wiedemann-Merdinoglu et al. (2006)			
<i>Plasmopara viticola</i>	<i>Rpv3</i>	18		UDV112	Regent	Lemberger	153	Regent		Welter et al. (2007)	Regent and Bianca descend from Seibel 4614 (= <i>Rpv3</i> <sup>299-279</sup> = <i>Rpv3-1</i> )		
			24.9	UDV305	Chardonnay	Bianca	116	Bianca		Bellin et al. (2009)			
			26.9	VMC7f2	Regent	RedGlobe	206	Regent		van Heerden et al. (2014)			
			24.9	UDV305				Seibel 4614	<i>V. rupestris</i>	Di Gaspero et al. (2012)	pedigree analysis		
<i>Rpv3.1</i> (= <i>Rpv3</i> <sup>299-279</sup> )			26.1	UDV737					<i>V. rupestris</i>	Zyprian et al. (2016)			
			25.9	GF18-06	GF18-08	GF.GA-47-42	Villard blanc	151	Villard blanc				



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<i>Plasmopara viticola</i>	<i>Rpv27</i>	18	24.6	VVCS1H077H16R1-1	Norton	Cabernet Sauvignon	182	Norton	<i>V. aestivalis</i>	Sapkota et al. (2019)	
			26.0	UDV737							
<i>Plasmopara viticola</i>	<i>Rpv28.1</i>	10	1.3	S10_1285522	V. rupestris B38	V. riparia HP-1	136	V. rupestris B38	<i>V. rupestris</i>	Bhattarai et al. (2021)	physical position based on 12X.v2
			3.0	S10_3024940							
<i>Plasmopara viticola</i>	<i>Rpv29</i>	14	21.6	chr14_21613512_C_T	Mgaloblishvili	Mgaloblishvili	132	Mgaloblishvili	<i>V. vinifera</i>	Sargolzaei et al. (2020)	84 OPs of Mgaloblishvili, 48 genotypes of Georgian germplasm collection
<i>Plasmopara viticola</i>	<i>Rpv30</i>	3	16.2	cn_C_T_chr3_16229046	Mgaloblishvili	Mgaloblishvili	132	Mgaloblishvili	<i>V. vinifera</i>	Sargolzaei et al. (2020)	84 OPs of Mgaloblishvili, 48 genotypes of Georgian germplasm collection
<i>Plasmopara viticola</i>	<i>Rpv31</i>	16	21.3	li_T_C_chr16_21398409	Mgaloblishvili	Mgaloblishvili	132	Mgaloblishvili	<i>V. vinifera</i>	Sargolzaei et al. (2020)	84 OPs of Mgaloblishvili, 48 genotypes of Georgian germplasm collection
<i>Plasmopara viticola</i>	<i>Rpv32</i>									Malagol et al. (in preparation)	
<i>Plasmopara viticola</i>	<i>Rpv33</i>	9	0.9	rh_chr9_893002	<i>Vitis x doaniana</i>	Chardonnay	220	<i>V. acerifolia</i>	<i>V. acerifolia</i>	Zou et al. (2023)	<i>Vitis x doaniana</i> (= <i>V. mustangensis</i> x <i>V. acerifolia</i> )
<i>Plasmopara viticola</i>			1.3	rh_chr9_1305204							
<i>Plasmopara viticola</i>	<i>Rpv34</i>									Cadle-Davidson et al. (in preparation)	
<i>Plasmopara viticola</i>	<i>Rpv35</i>									Cadle-Davidson et al. (in preparation)	
<i>Plasmopara viticola</i>	<i>Rpv36</i>									De Lorenzis et al. (in preparation)	
<i>Plasmopara viticola</i>	<i>Rpv37</i>									De Lorenzis et al. (in preparation)	
<i>Xiphinema index</i>	<i>XiR1</i>	19	20.9	VMC5a10	<i>V. rupestris</i>	<i>V. arizonica</i>	185	<i>V. arizonica</i>	<i>V. arizonica</i>	Xu et al. (2008)	
			20.9	1N2R3b						Hwang et al. (2010)	
				M4F3R							
<i>Xiphinema index</i>	<i>XiR2</i>	9		VVBX-A-06	VRH8771	Cabernet Sauvignon	135	VRH8771		Rubio et al. (2020)	
<i>Xiphinema index</i>	<i>XiR3</i>	10		SC8-03	VRH8771	Cabernet Sauvignon	135	VRH8771		Rubio et al. (2020)	
<i>Xiphinema index</i>	<i>XiR4</i>	18	29.1	UDV108	VRH8771	Cabernet Sauvignon	135	VRH8771		Rubio et al. (2020)	
Morphology											
Berry size (berry weight)	<i>Be size</i>	18	25.9	SCC8	MTP2223-27	MTP2121-30	139		<i>V. vinifera</i>	Doligez et al. (2002)	Only one major QTL for berry size is indicated. There are several other QTLs described in the literature.
			26.9	VMC7f2	Dominga	Autumn Seedless	118			Cabezas et al. (2006)	
					Ruby Seedless	Thompson Seedless	144			Mejia et al. (2007); Mejia et al. (2011)	
					Italia	Big Perlon	163			Costantini et al. (2008)	
Fleshless berry	<i>Flb</i>	18	0.9	VMC2a3	Chardonnay	Ugni Blanc Mutant	71	Ugni Blanc	<i>V. vinifera</i>	Fernandez et al. (2006)	Mutant
GA insensitive dwarf mutant	<i>Vgai1</i>	1	4.9					Pinot Meunier		Boss & Thomas (2002)	periclinal chimera mutant
Leaf hairs	<i>LH1</i>	5	0.9	Nifts5-50363	Muscat of Alexandria	Campbell Early	95	Muscat of Alexandria	<i>V. vinifera</i>	Kono et al. (2018)	reducing leaf hair density; confers DM susceptibility
Leaf variegation	<i>Lvar1</i>	14	26.1	14_26071477	MN1220	MN1326	119			Olson et al. (2022)	locus validated with 2 other populations and with GWAS
			28.7	14_28696427							
Leaf variegation	<i>Lvar2</i>	11	16.3	11_16272242	MN1256	MN1327	57			Olson et al. (2022)	
			18.4	11_18433707							
Seed development inhibitor (Seedlessness)	<i>SdI</i>	18	25.9	SCC8	MTP2223-27	MTP2121-30	139			Doligez et al. (2002)	
			23.2	VMC6f11	Dominga	Autumn Seedless	118	Autumn Seedless		Cabezas et al. (2006)	
			26.9	VMC7f2							
			26.9	VMC7f2	Italia	Big Perlon	163	Big Perlon		Costantini et al. (2008)	
			26.9	p3_VvAGL11	Ruby Seedless	Sultana	139			Mejia et al. (2011)	
Sex	<i>Sex</i>	2	3.7	VVMD34	Horizon	Illinois 547-1	58			Dalbó et al. (2000)	
			4.2	VVS3	Ramsey	Riparia Gloire	188			Lowe and Walker (2006)	
			4.9	VVlb23	<i>V. rupestris</i>	<i>V. arizonica</i>	181			Riaz et al. (2006)	
			5.0	APT3Indel	V3125	Börner	202			Fechter et al. (2012)	
			4.7	SNP4C_1	Moscato Bianco	<i>V. riparia</i> WR63	340			Battilana et al. (2013)	
			4.9	VVlb23	Muscat Ottonel	Malvasia aromatica di	91				
			4.9	VSVV007						Piqq et al. (2014)	
			5.0	VSVV010							

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<b>Phenology</b>											
Berry skin color	<i>BeCo</i>	2	8.2	VMC5g7	MTP3140	MTP2223-27	139			Doligez et al. (2002)	
			17.5	VMC8c2						Kobayashi et al. (2004)	
			14.2	MybA1							
Véraison	<i>Ver</i>	16	13.7	VMCle11	Regent	Lemberger	153	Regent		Fischer et al. (2004)	For véraison (begin of ripening) several QTLs are published. This data here is incomplete.
										Costantini et al. (2008)	
Véraison	<i>VerI</i>	16	15.8	UDV052	GF.GA-47-42	Villard blanc	151	GF.GA-47-42		Zyprian et al. (2016)	
				SNP1092P11R						Zyprian et al. (2016)	
Véraison	<i>Ver2</i>	18		SPS_P_SNP632GF	GF.GA-47-42	Villard blanc	151				
Growth cessation [photoperiod-induced]		13	8.1	VMC3D12	(V. <i>riparia</i> x Seyval)	self-pollinated	113			Garris et al. (2009)	greenhouse
			27.0	VVIp10							field
			9.1	VMC6G1							
		11	19.1	VVIp36c			95				
<b>Metabolites</b>											
Anthocyanin 3-monoglycosides	<i>Ufgt</i>	16	2.3	UFGT	Regent	Lemberger	153			Fischer et al. (2004)	
Anthocyanin 3,5-diglycosides	5-GT	9	6.5		Regent	Lemberger	153	Regent		Hausmann et al. (2009)	
										Janvary et al. (2009)	
										Yang et al. (2014)	
Monoterpene content	<i>Mtc</i>	5	3.8	DXS1	Italia	Big Perlon	163		<i>V. vinifera</i>	Battilana et al. (2009)	
Muscat flavor		1	6.1	VVIq57	MTP2687-85 (Olivette noire x Ribol)	Muscat of Hamburg	174	Muscat of Hamburg	<i>V. vinifera</i>	Doligez et al. (2006)	
			5	VVMD27							
			5.7	VrZAG79							
			7	22.6							
Geraniol content		5	4.5	VVMD27	MTP2687-85 (Olivette noire x Ribol)	Muscat of Hamburg	174	Muscat of Hamburg	<i>V. vinifera</i>	Doligez et al. (2006)	
			5.7	VrZAG79							
			13	22.5							
			16	22.0							
Linalool content	<i>Lin</i>	2	4.8	VVlb23	MTP2687-85 (Olivette noire x Ribol)	Muscat of Hamburg	174	MTP2687-85 (Olivette noire x Ribol)	<i>V. vinifera</i>	Doligez et al. (2006)	
			7.0	VMC2C10.1							
		5	4.5	VVMD27							
			5.7	VrZAG79							
Linalool content	<i>Lin</i>	10	cnd41	Italia	Big Perlon	V. riparia WR63	174		<i>V. vinifera</i>	Battilana et al. (2009)	
			1.2	VVIh01							
			1.4	VrZAG67							
			1.3	VrZAG64							
Linalool content	<i>Lin</i>	10	1.1	VMC3d7	Muscat Ottonel	Gewuerztraminer	121		<i>V. vinifera</i>	Duchene et al. (2009)	
			1.1	VMC3d7							
			22.3	VVMD5							
			22.3	VVMD5							
Nerol content		5	4.5	VVMD27	MTP2687-85 (Olivette noire x Ribol)	Muscat of Hamburg	174	Muscat of Hamburg	<i>V. vinifera</i>	Doligez et al. (2006)	
			5.7	VrZAG79							
Isobutyl-methoxypyrazine (IBMP)	<i>VvOMT3</i>	3	2.2	VvOMT3	(Cabernet Sauvignon x Pinot Meunier)	self pollinated	64	Cabernet Sauvignon		Dunlevy et al. (2013)	3 significant QTLs for IBMP content
Malic acid concentration	<i>MA</i>	6	8.0		16-9-2	self pollinated	63			Yang et al. (2016)	16-9-2 = F1 of <i>V. riparia</i> x Seyval

Trait	Symbol	Chr	Position on chr [Mb]	Associated / flanking marker	Parent 1	Parent 2	Population size	Genotype of origin	Original species of trait	Reference	Comment
Iron deficiency [lime-induced chlorosis]		13	8.2	VMC3D12	Cabernet Sauvignon	Gloire de Montpellier	138	Gloire de Montpellier	<i>V. riparia</i>	Bert et al. (2013)	Only the major QTL is indicated. There are many other QTLs described.
			24.7	VMC2C7							
Mg [Mg content; chlorosis]		11	3.9	VVS2	Welschriesling	Sirius	92	Sirius		Mandl et al. (2006)	
			9.0	VMC6G1							
Yeast assimilable nitrogen	YAN	7	18.8		16-9-2	self-pollinated	63			Yang et al. (2016)	16-9-2 = F1 of <i>V. riparia</i> x Seyval